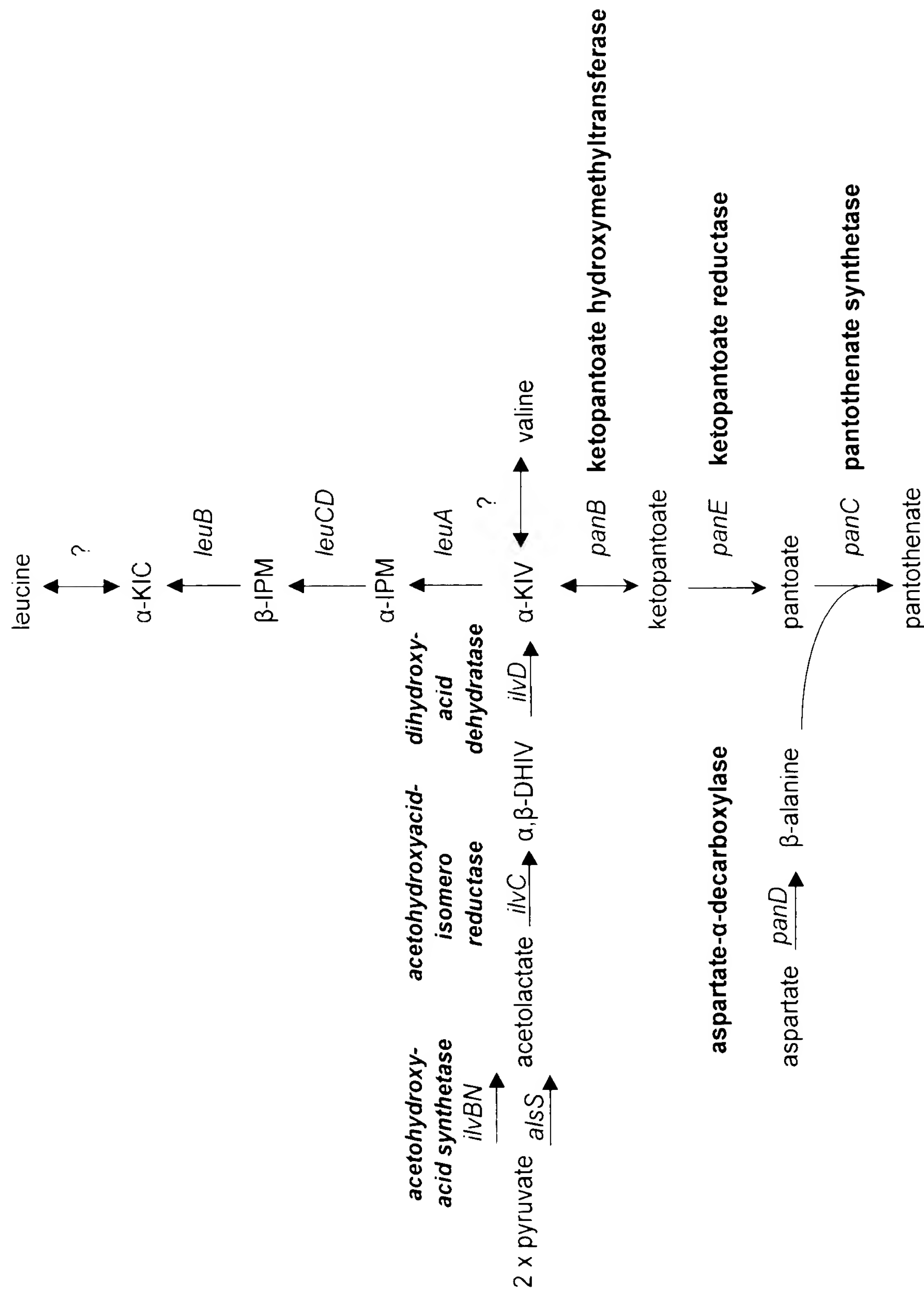


FIG.1



**Figure 2.** *Plasmid pAN240, containing sequences ligated upstream of the P<sub>26</sub>panBCD cassette, equivalent to the integrated version in strain PA221.*

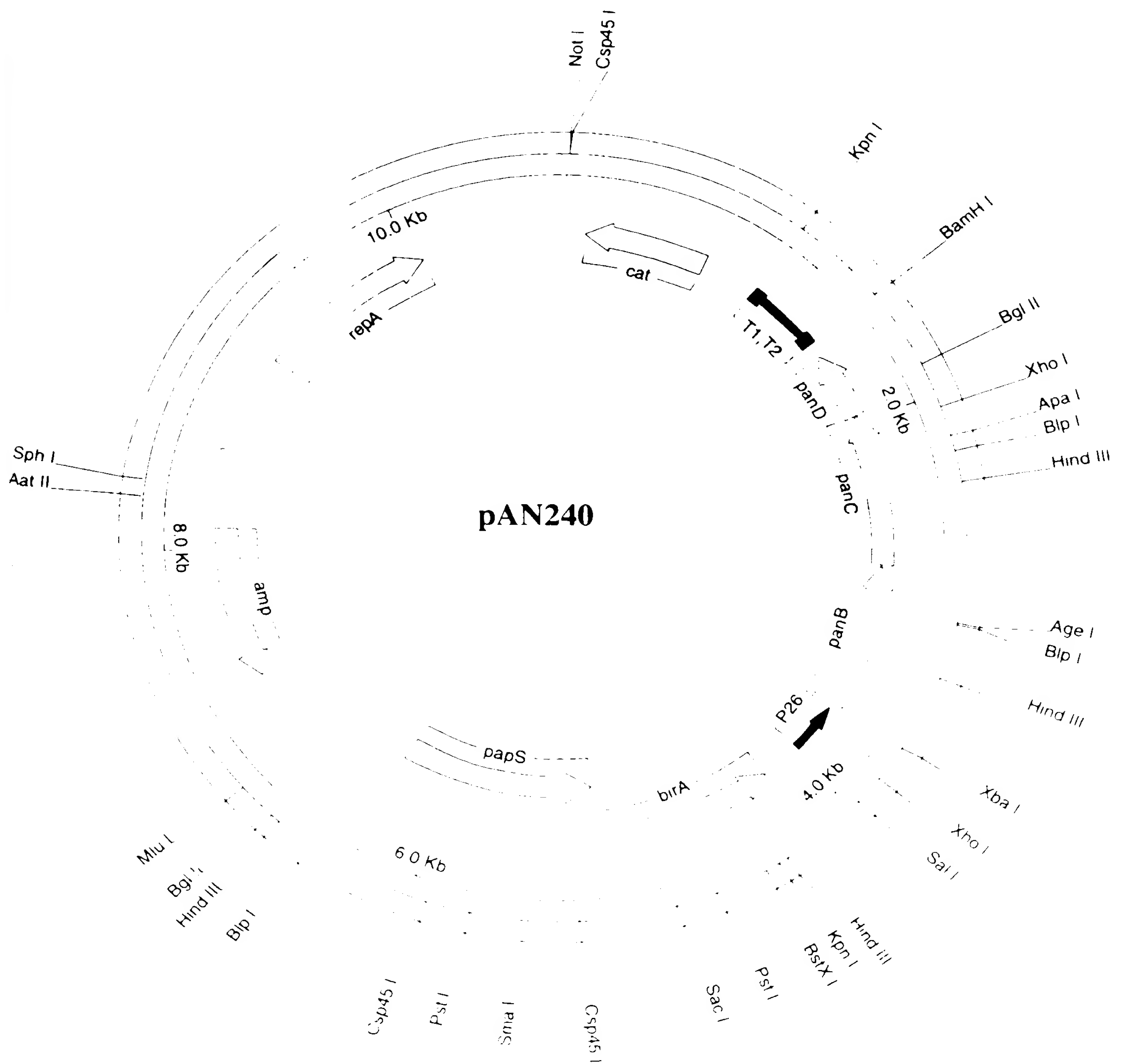


Figure 3A Plasmid pAN004, containing the panBCD operon expressed from P26 and RBS1.

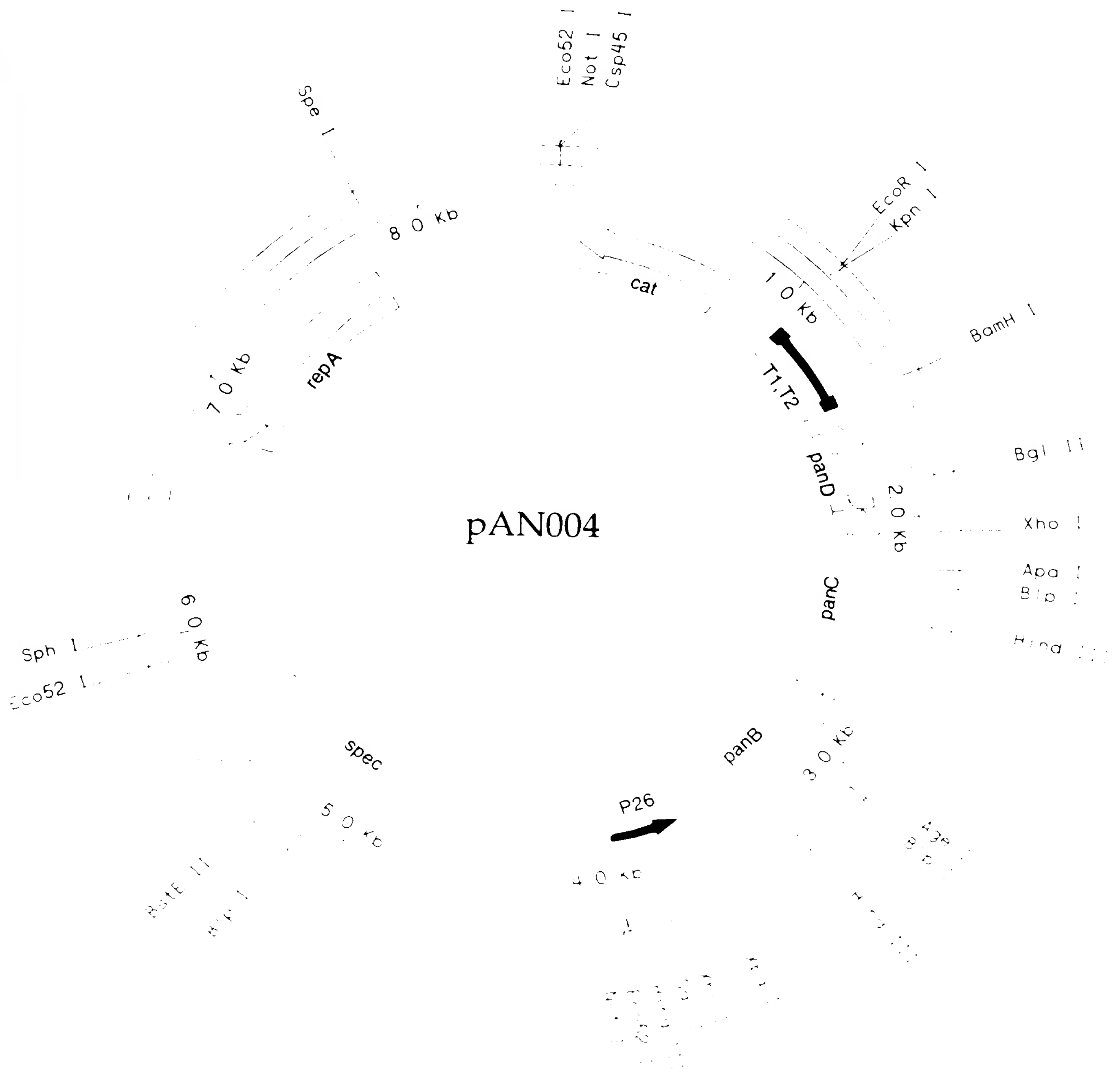


Figure 3 Plasmid pAN006, containing the panBCD operon expressed from P26 and RBS2.

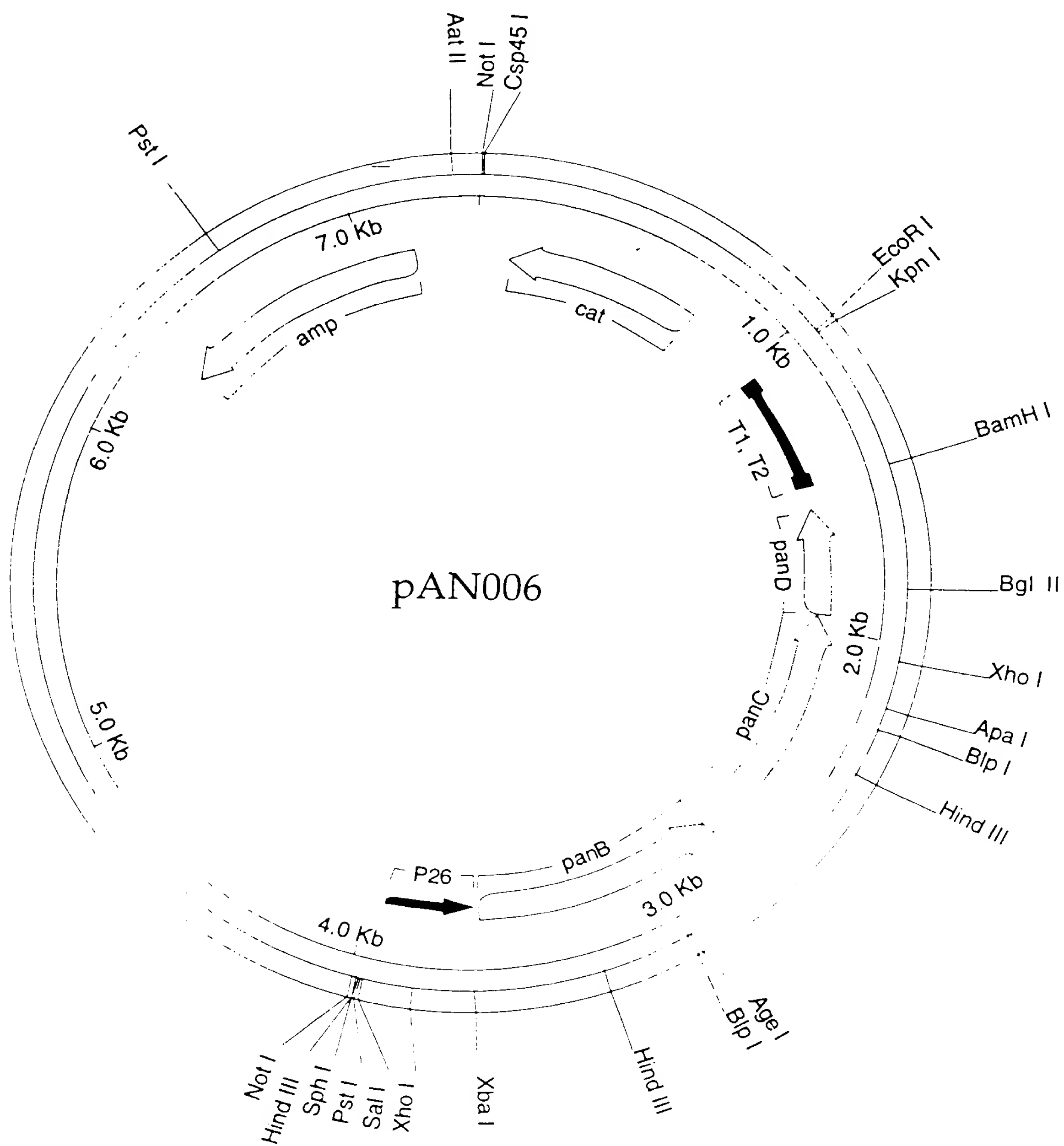


Figure 4 Plasmid pAN236, containing an integratable and amplifiable P26-RBS2-panE1 expression cassette.

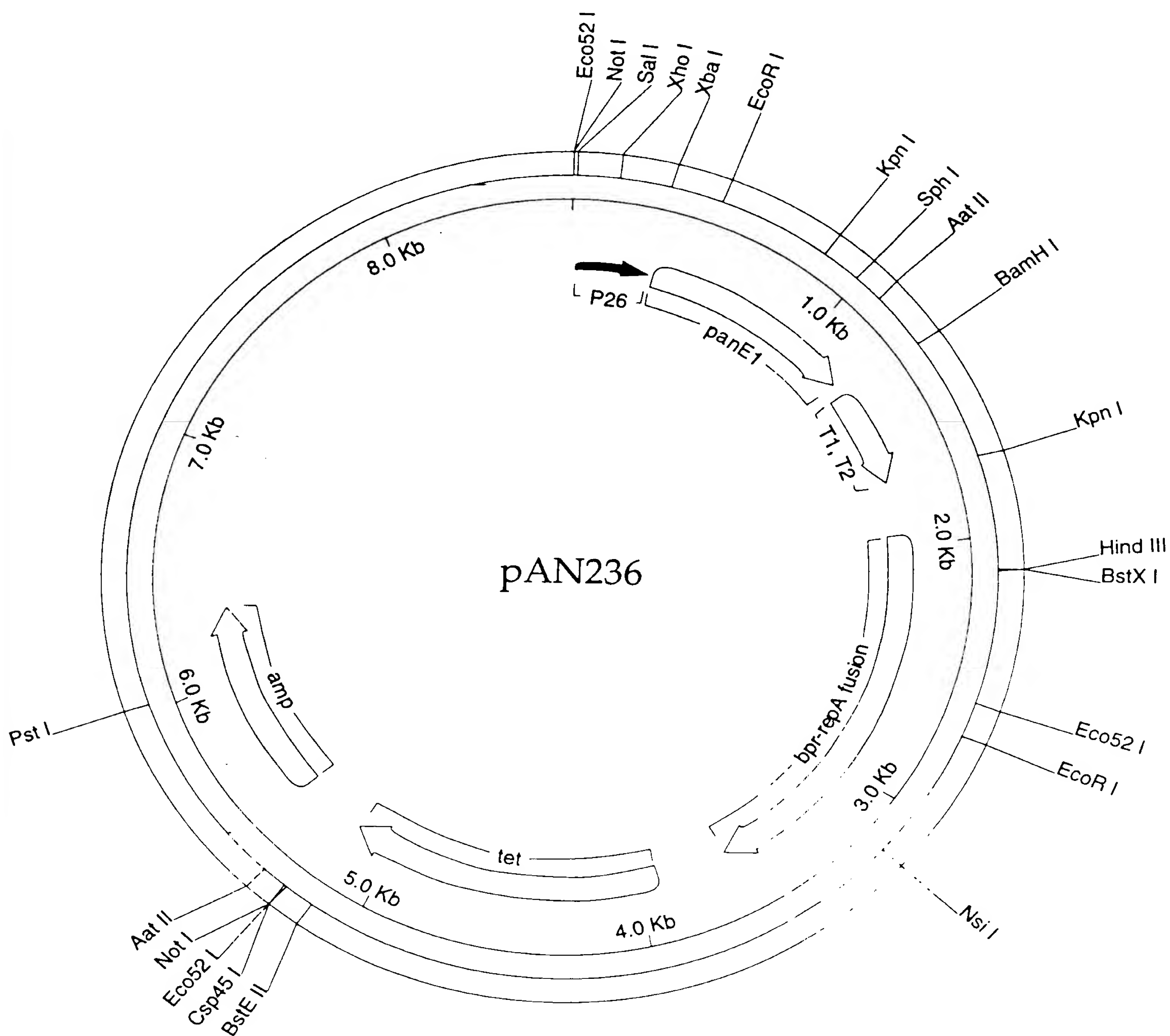


Figure 5 Construction of pAN423

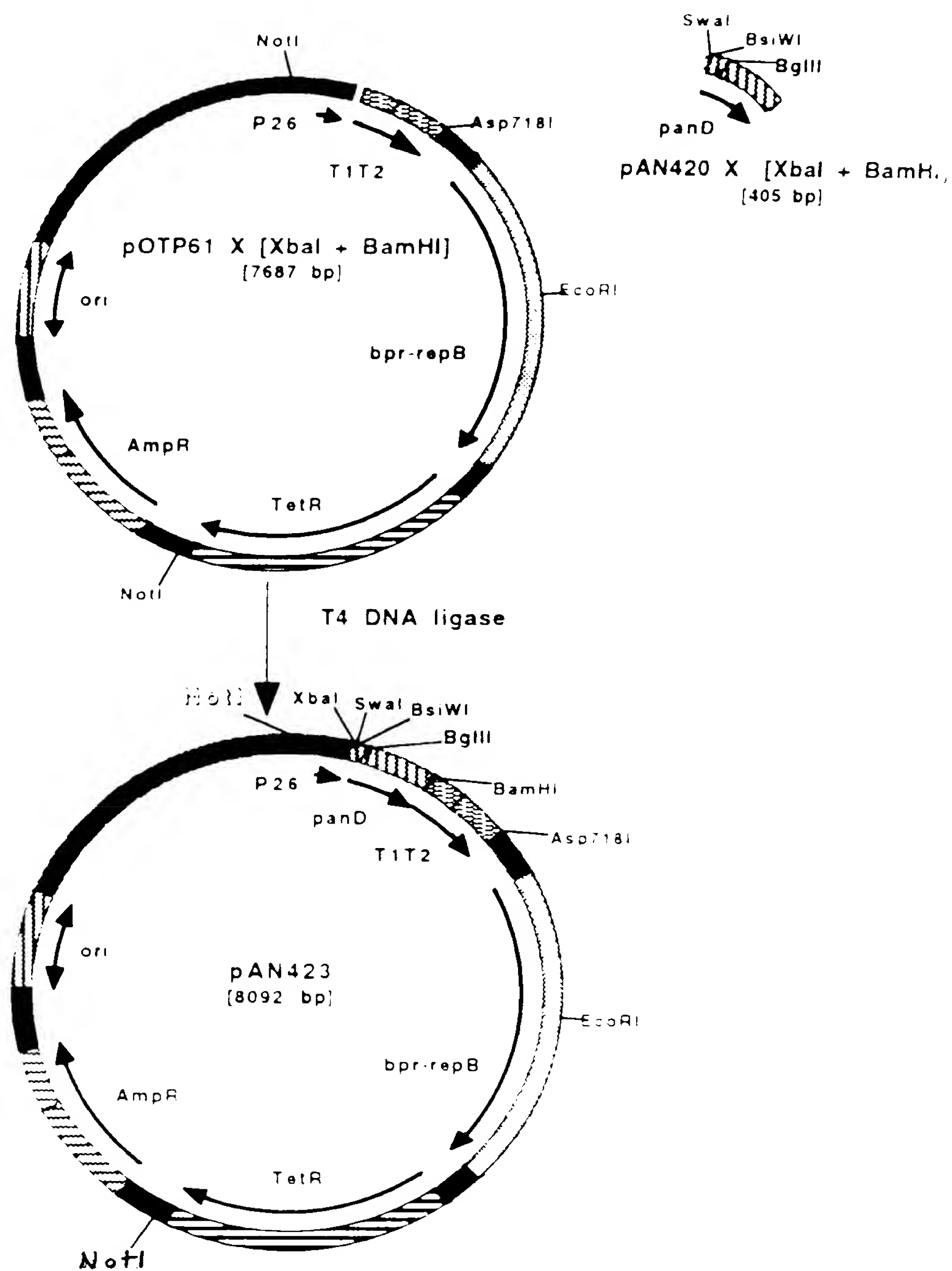
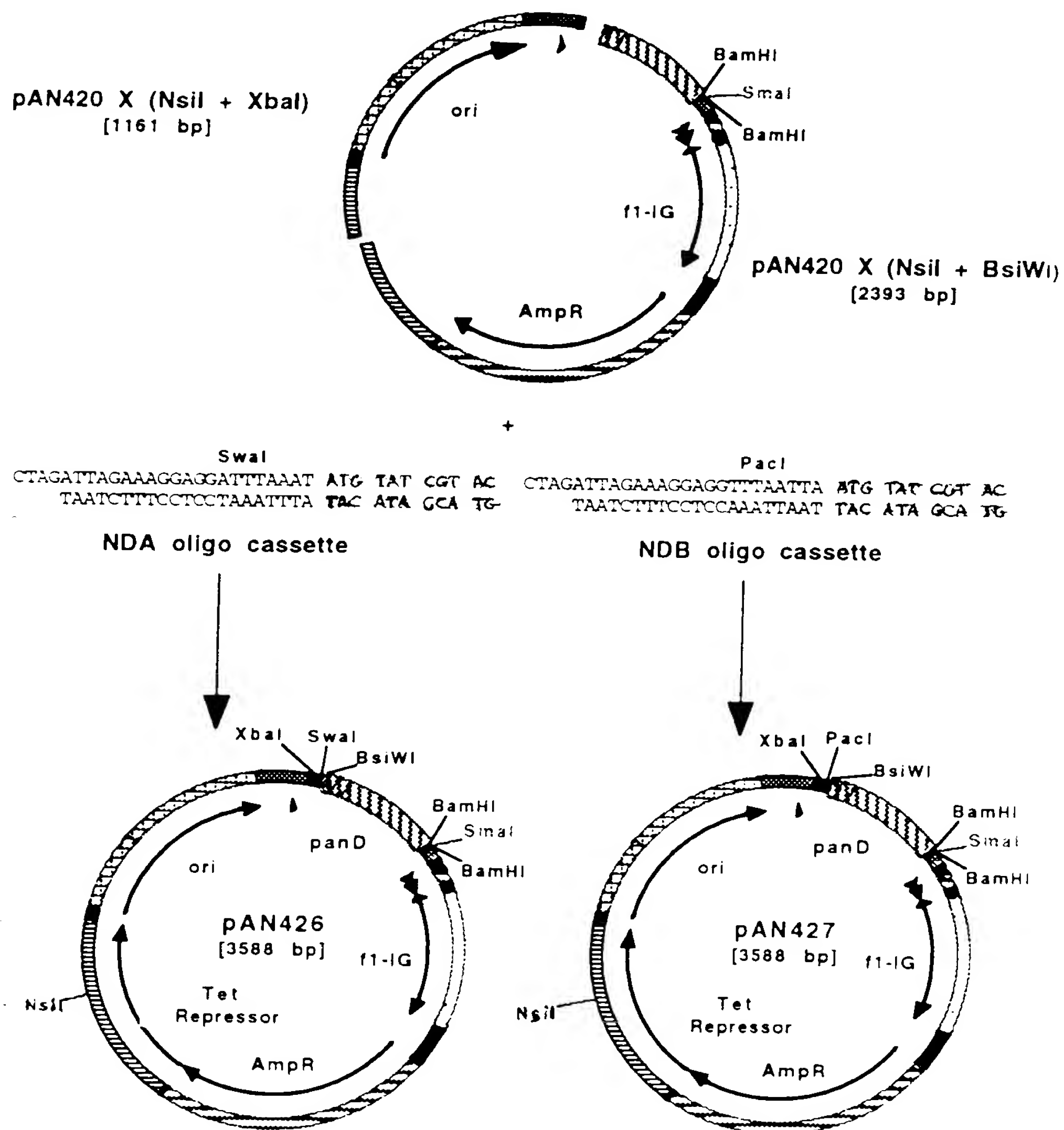


Figure 6 Construction of pAN426 and pAN427.



*Figure 7 Construction of pAN428 and pAN429.*

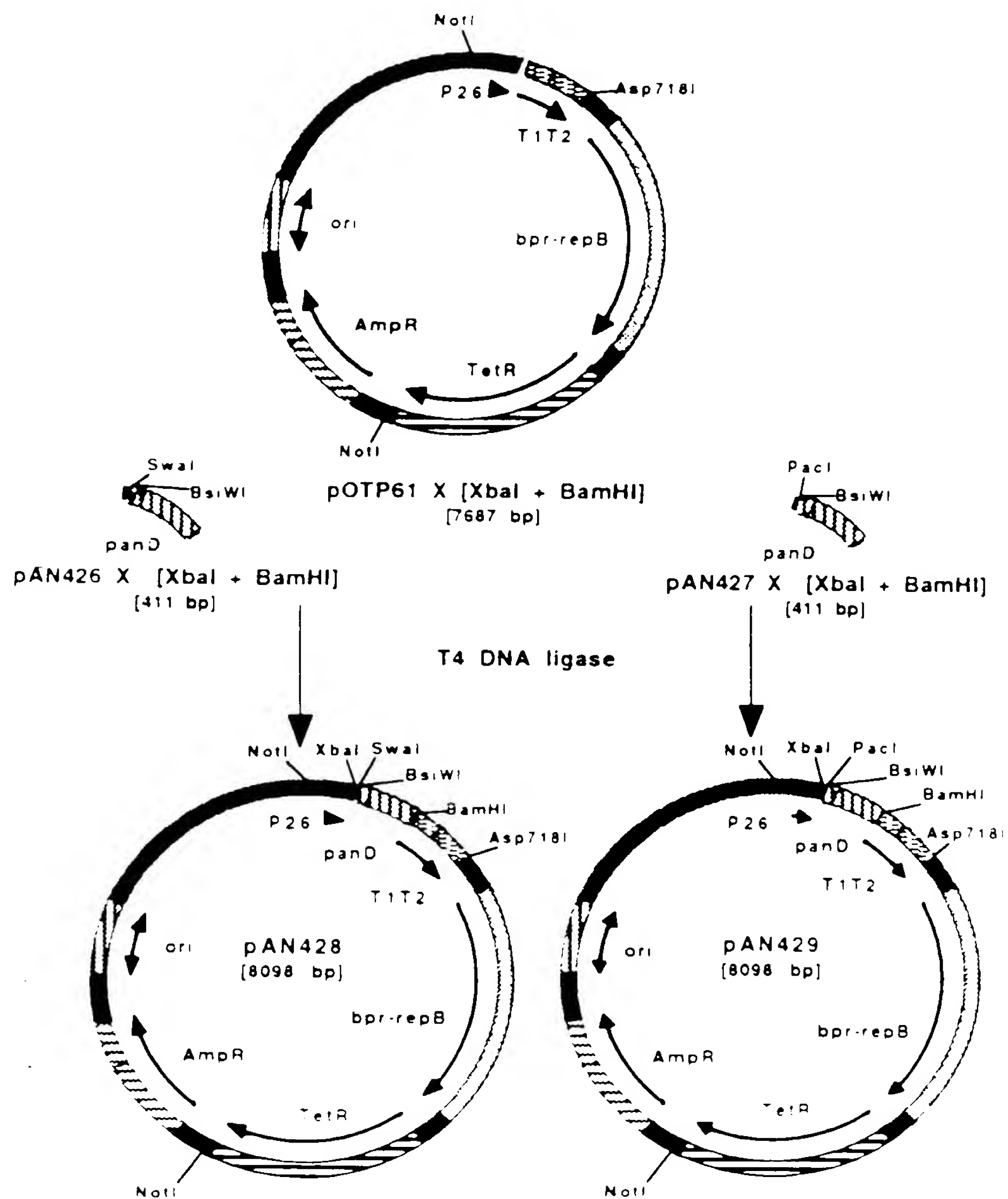




Figure 8. Construction of pAN431.

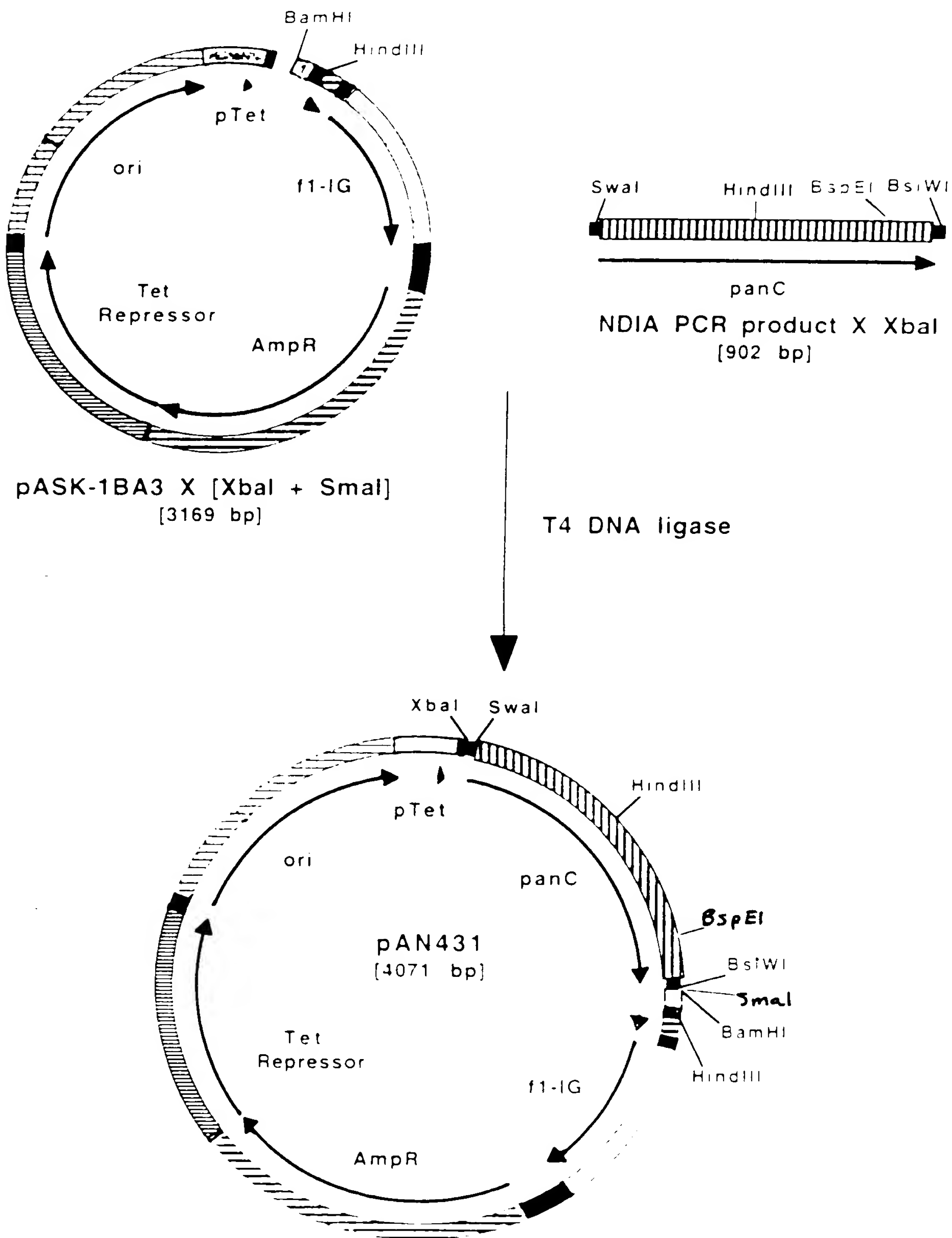


Figure 9. Construction of pAN441.

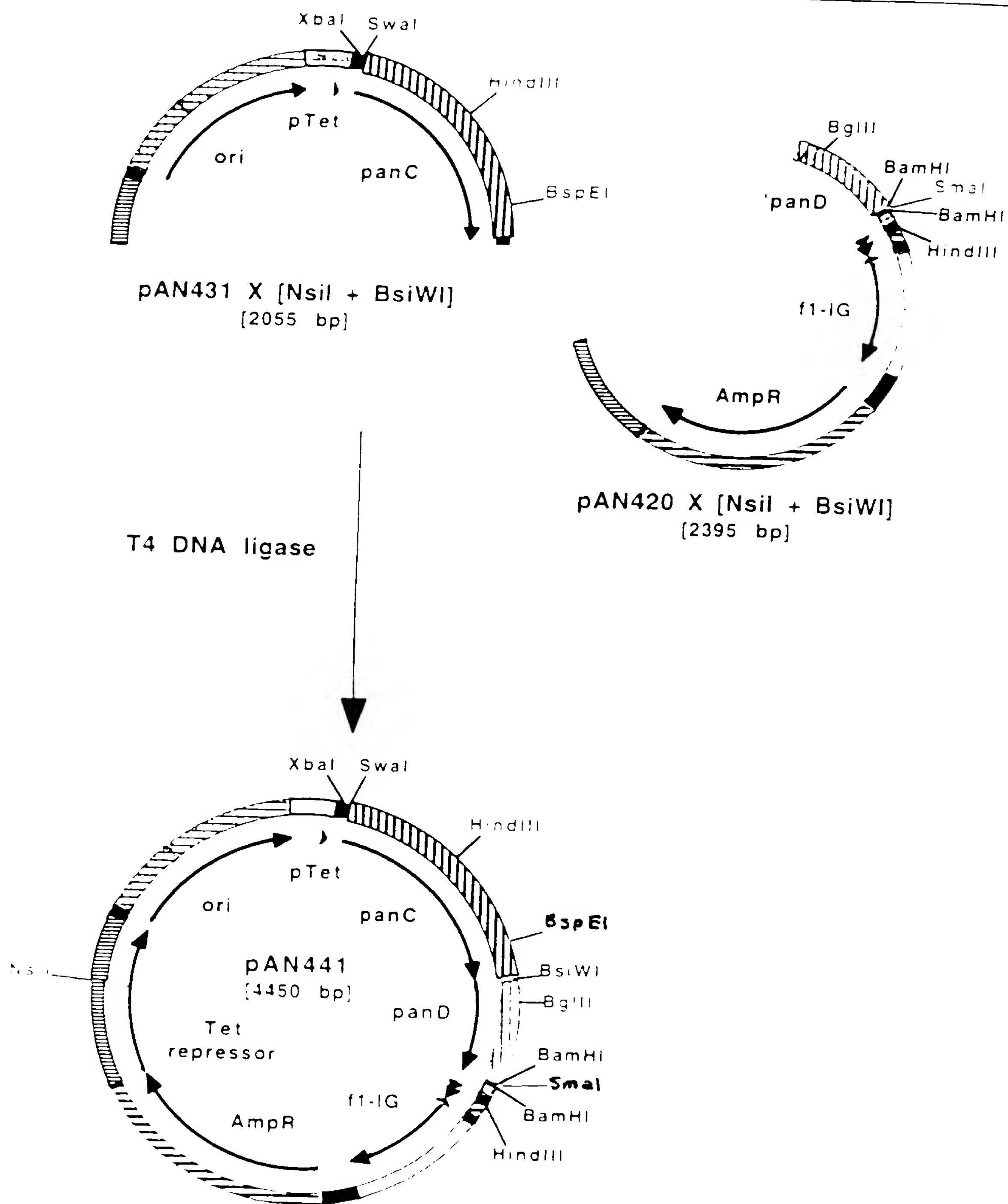


Figure 10. Construction of pAN440.

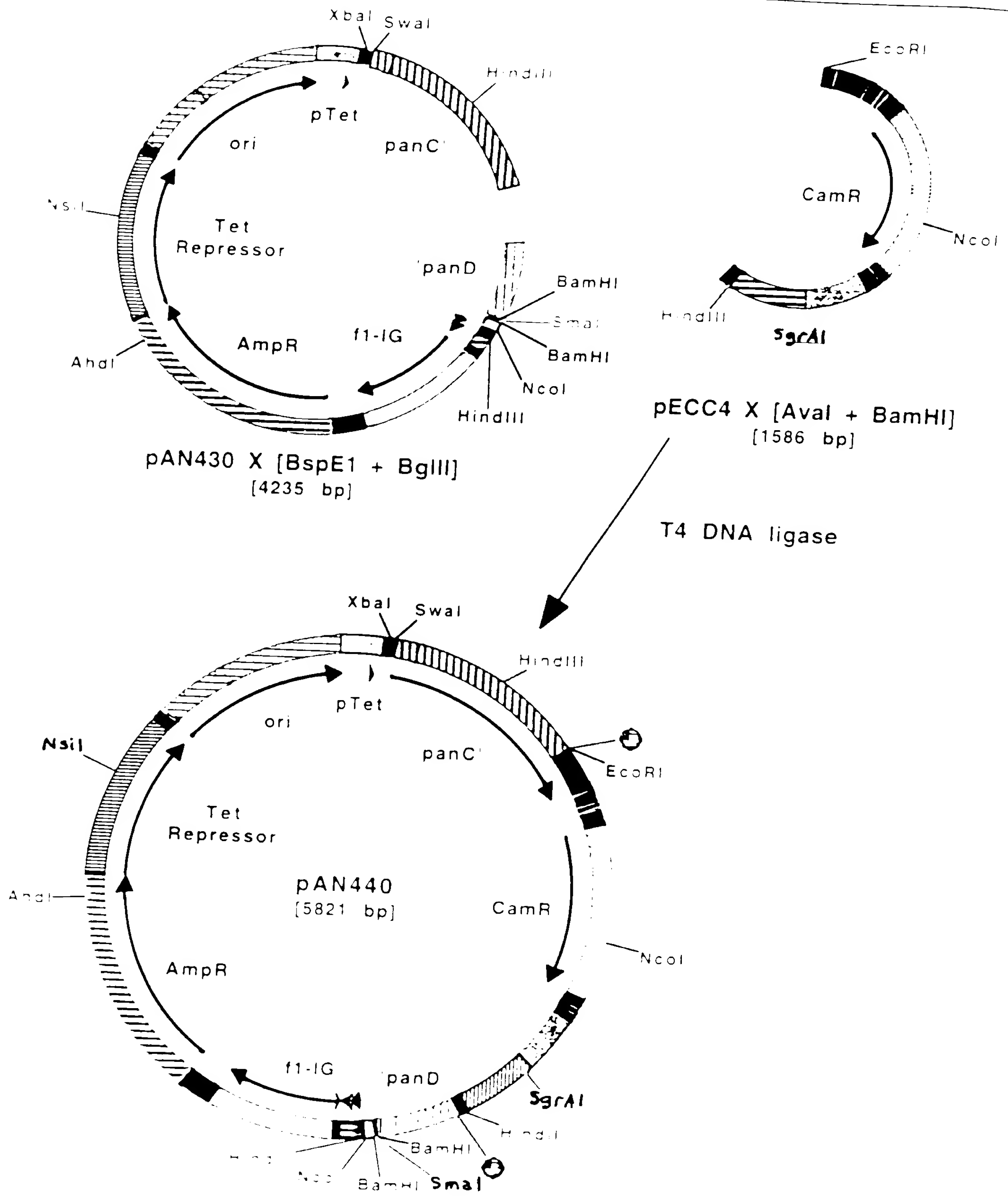


Figure 1| Structure of pAN251, a plasmid designed to integrate a single copy of P<sub>26</sub> panE1 at the panE1 locus by double crossover.

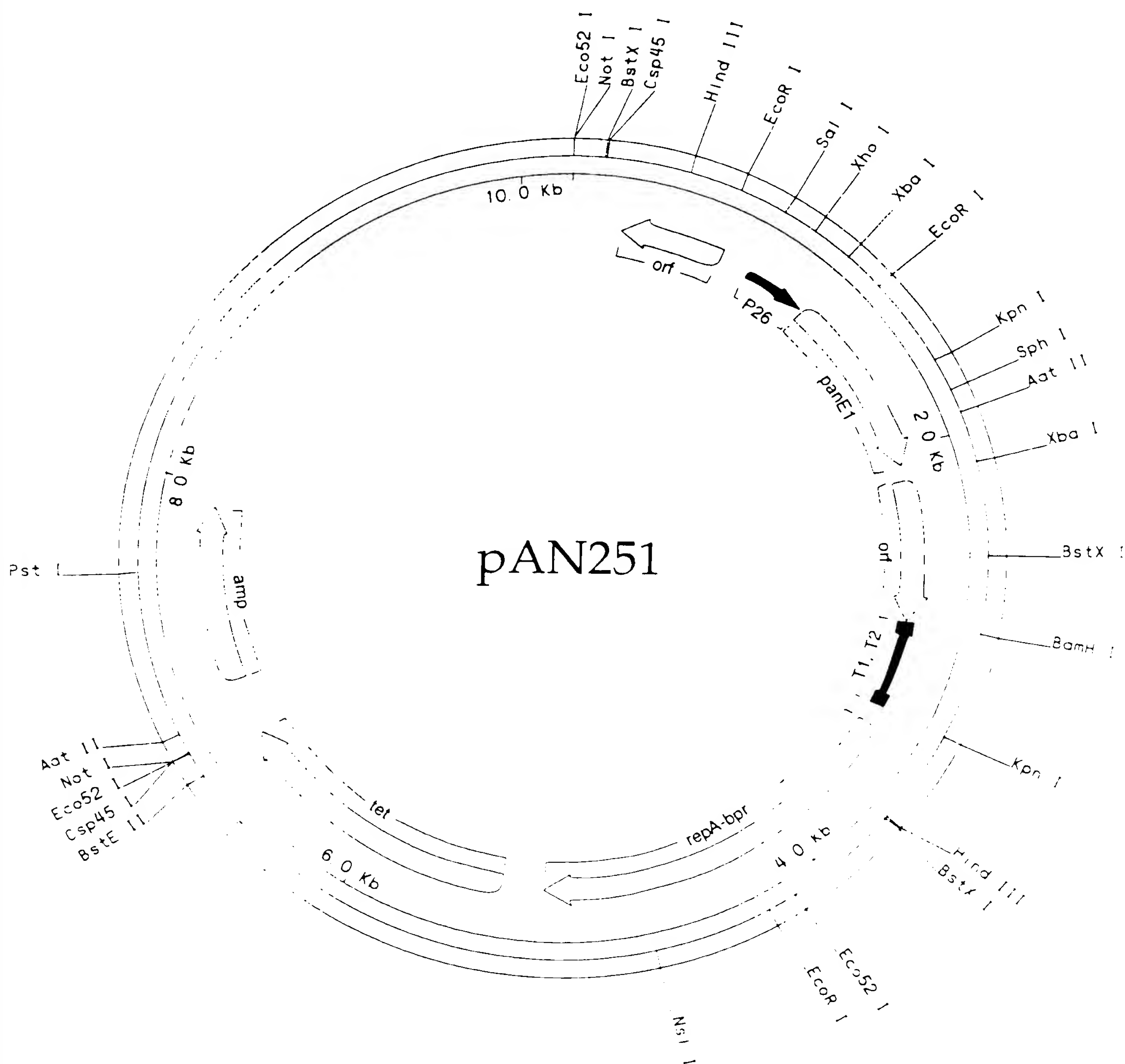


Figure 12 Structure of pAN267, a plasmid designed to stably integrate a P<sub>26</sub> ilvBNC cassette at the amyE locus.

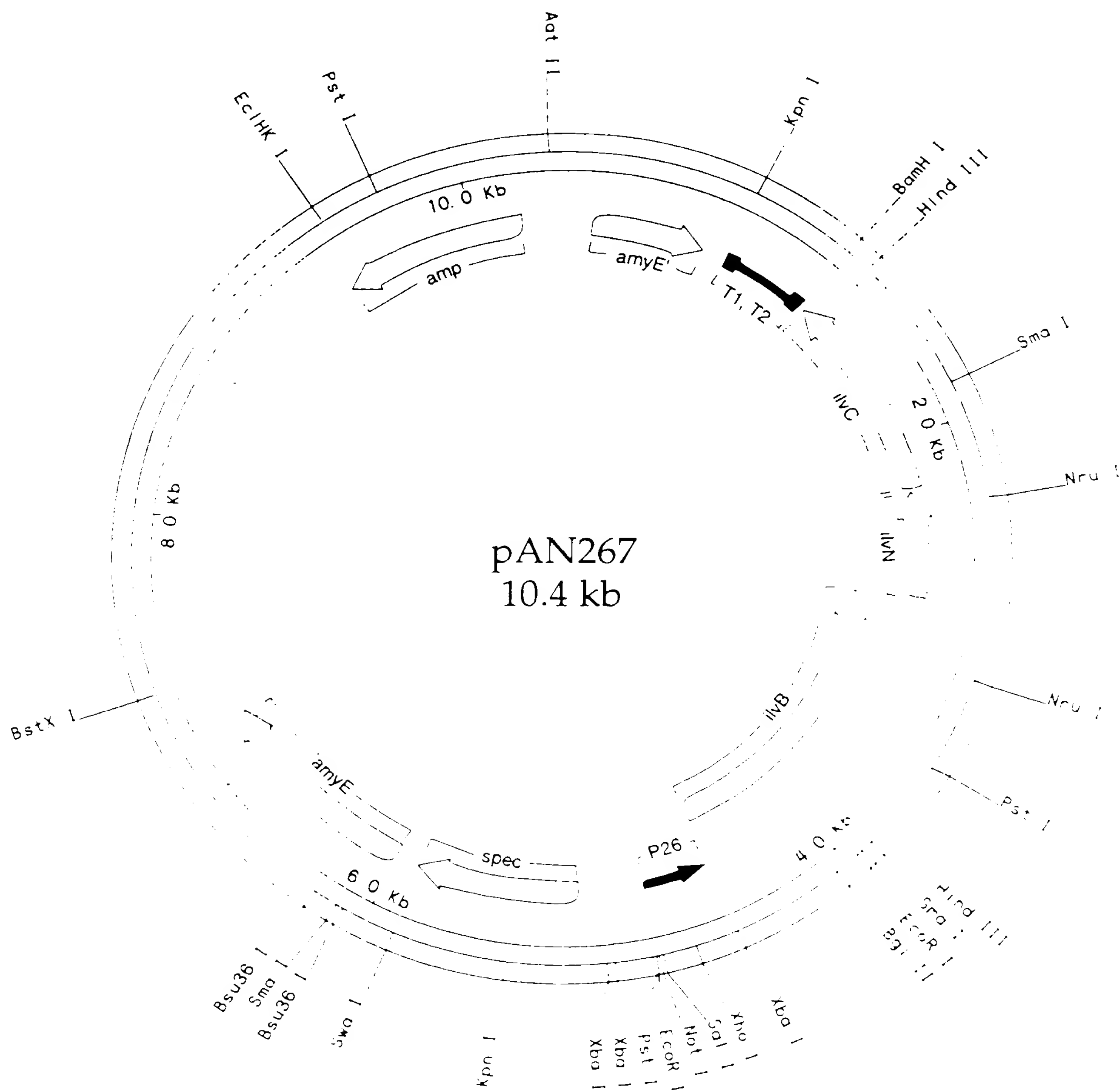


Figure 13 Structure of pAN257, a clone of *B. subtilis* *ilvD* in a low copy vector.

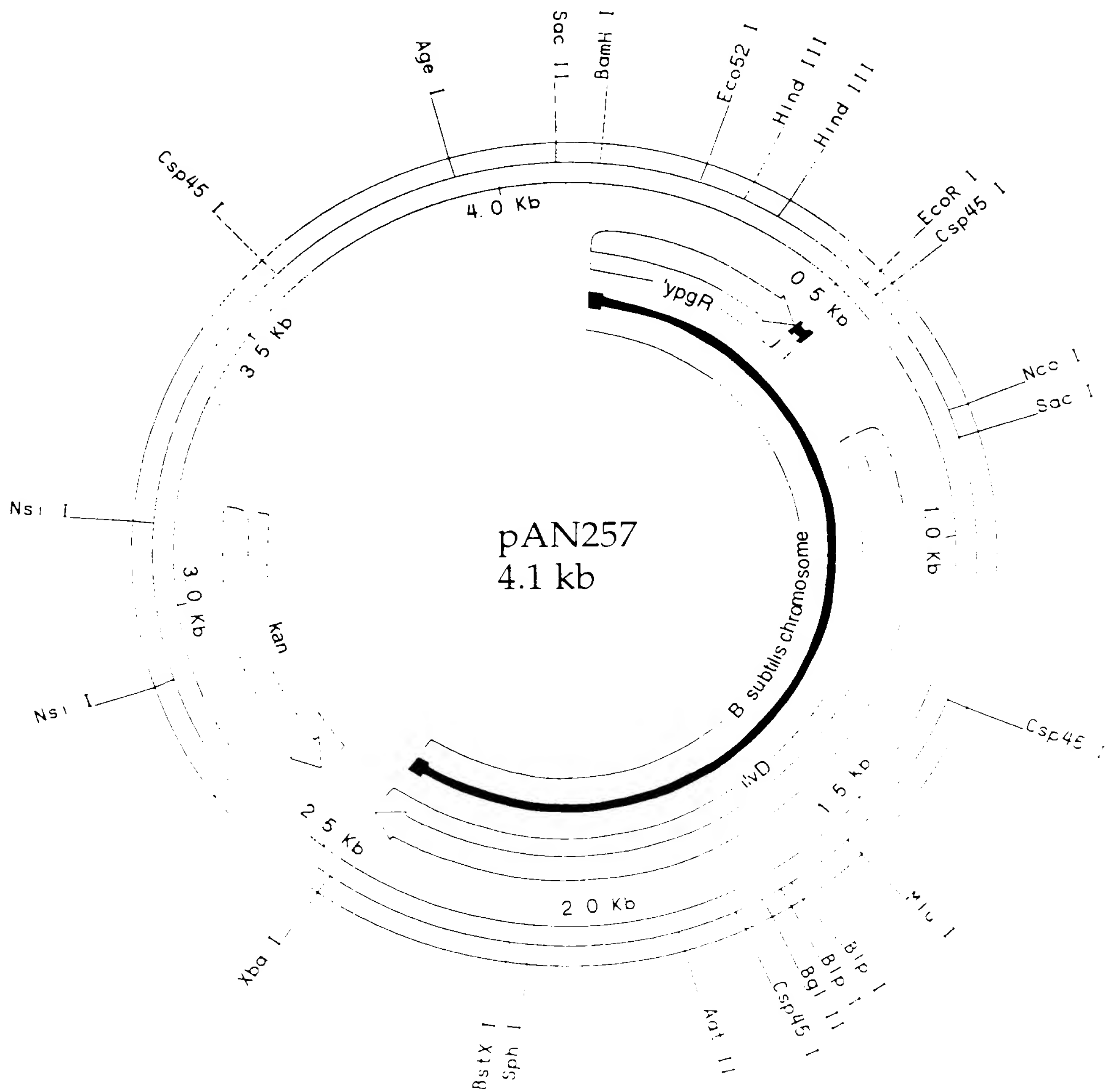


Figure 14 Structure of pAN263, designed to stably integrate a single copy of P<sub>26</sub> ilvD at the ilvD locus.

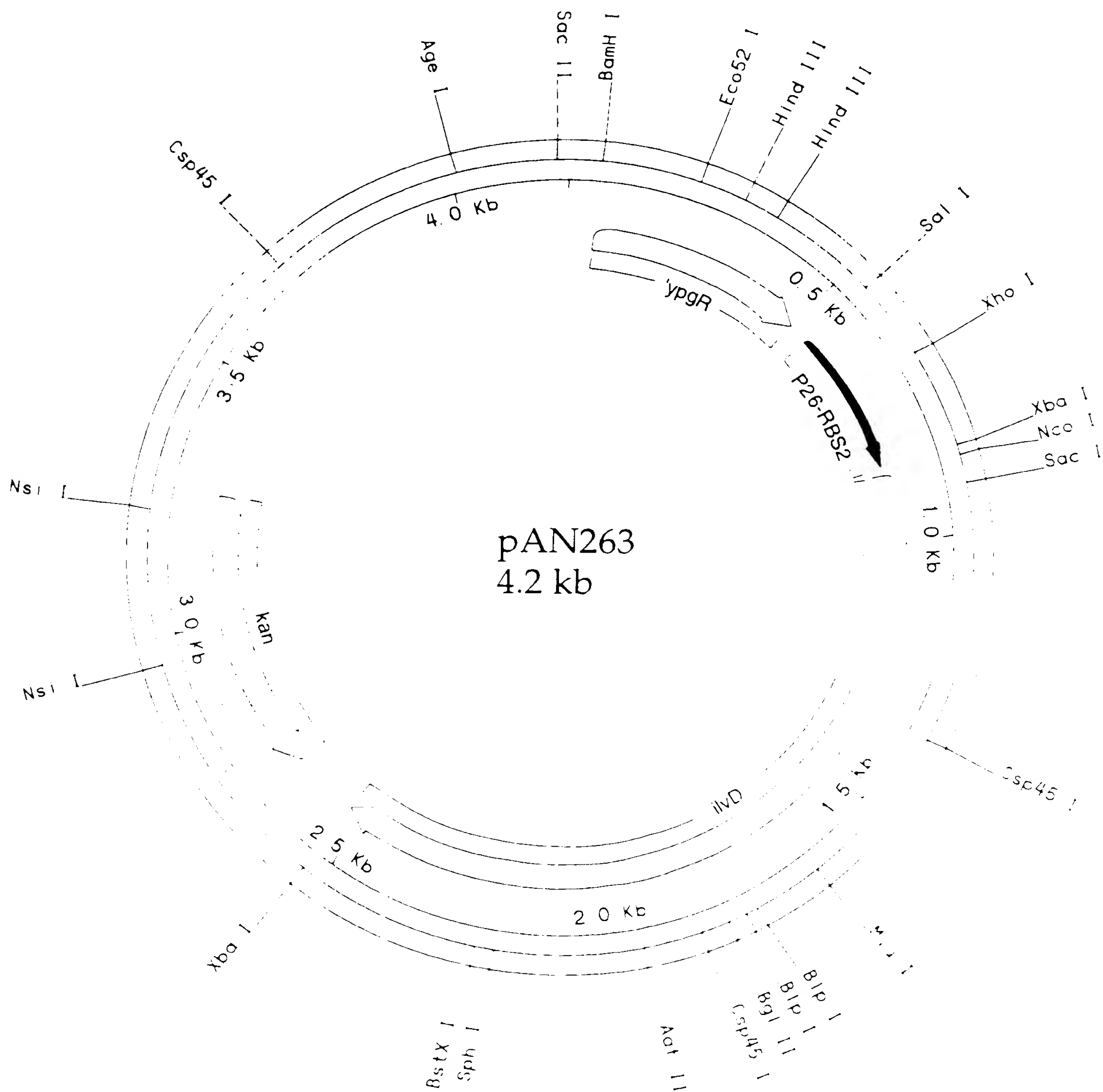


Figure 15 Structure of pAN261, designed to disrupt the *B. subtilis* *ilvD* gene with the *cat* gene.

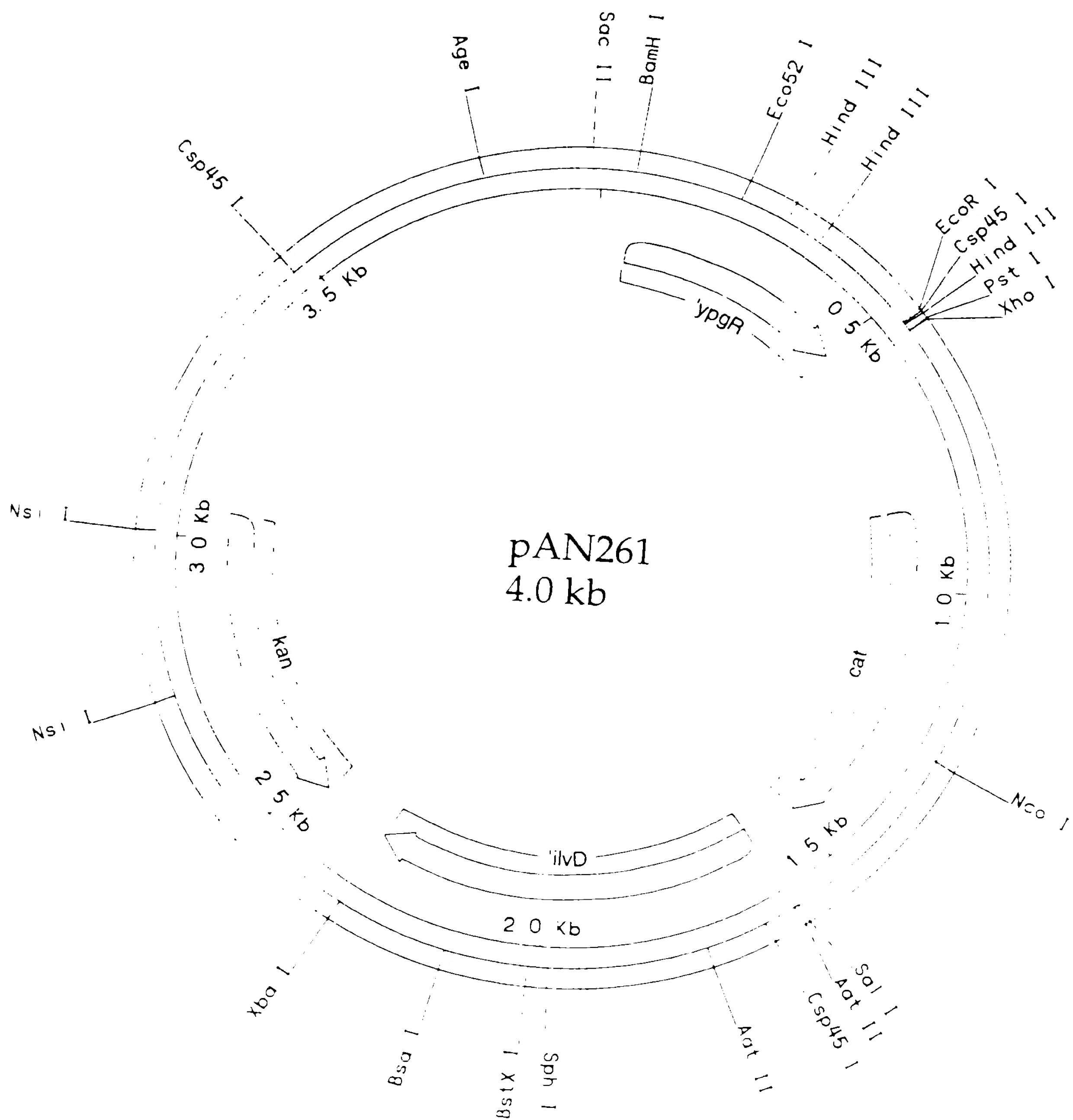




Figure 16

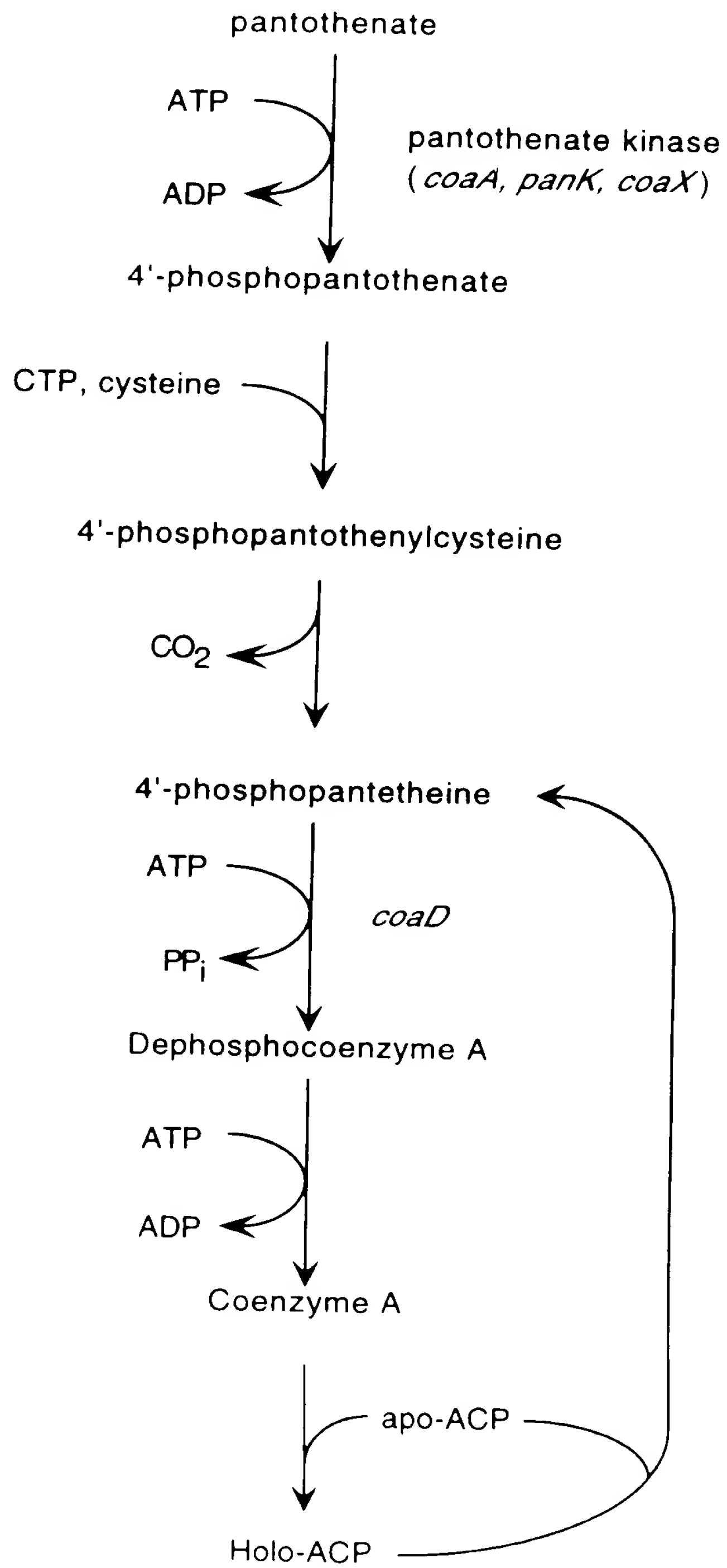


Figure 17 Structure of pAN296, designed to delete most of the *B. subtilis* *coaA* gene and substitute a chloramphenicol resistance gene.

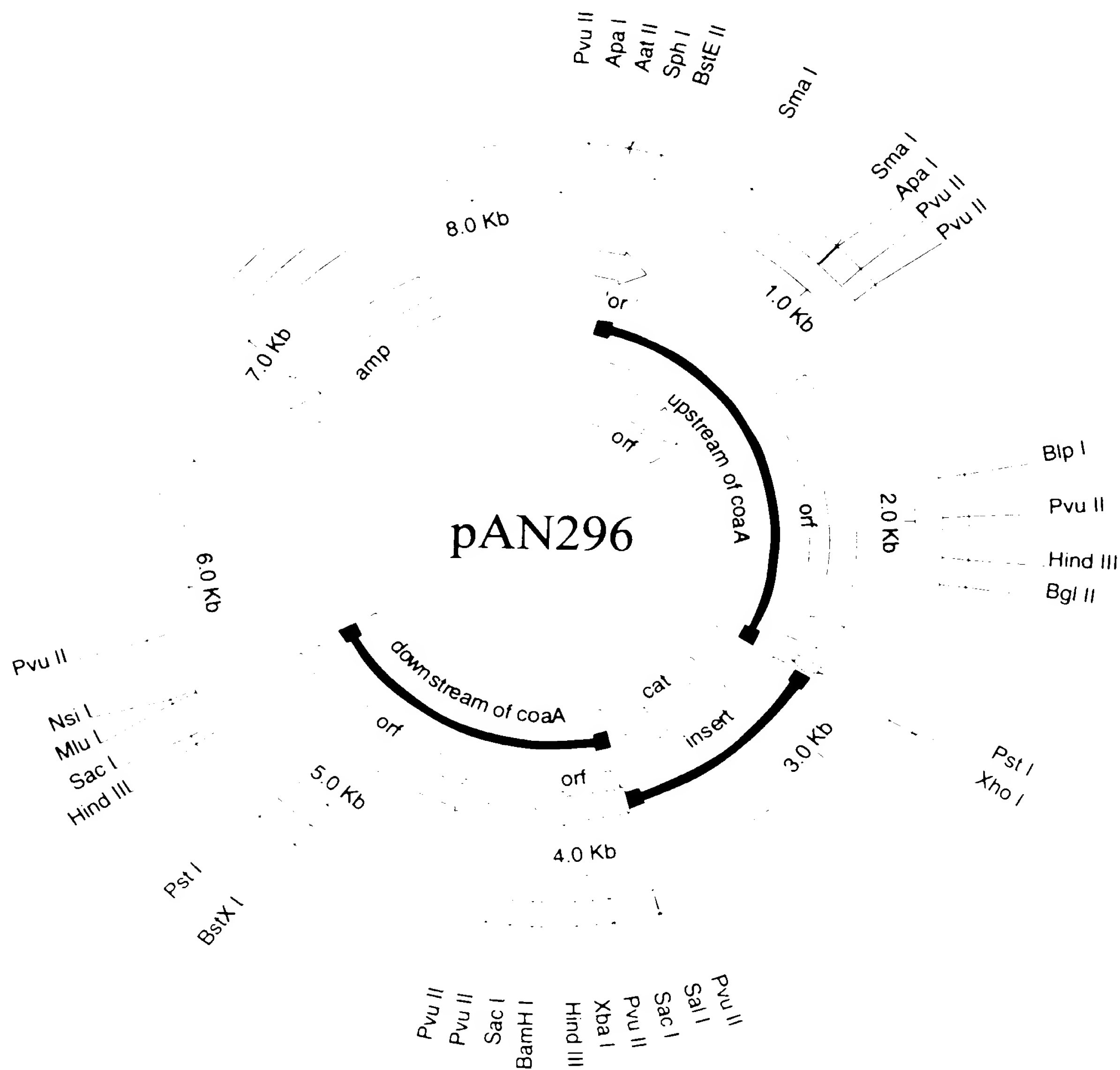


Figure 13 Structure of the *B. subtilis* chromosome in the region of the *coaA* gene. The scale is in base pairs and the significant open reading frames are shown by the open arrows.

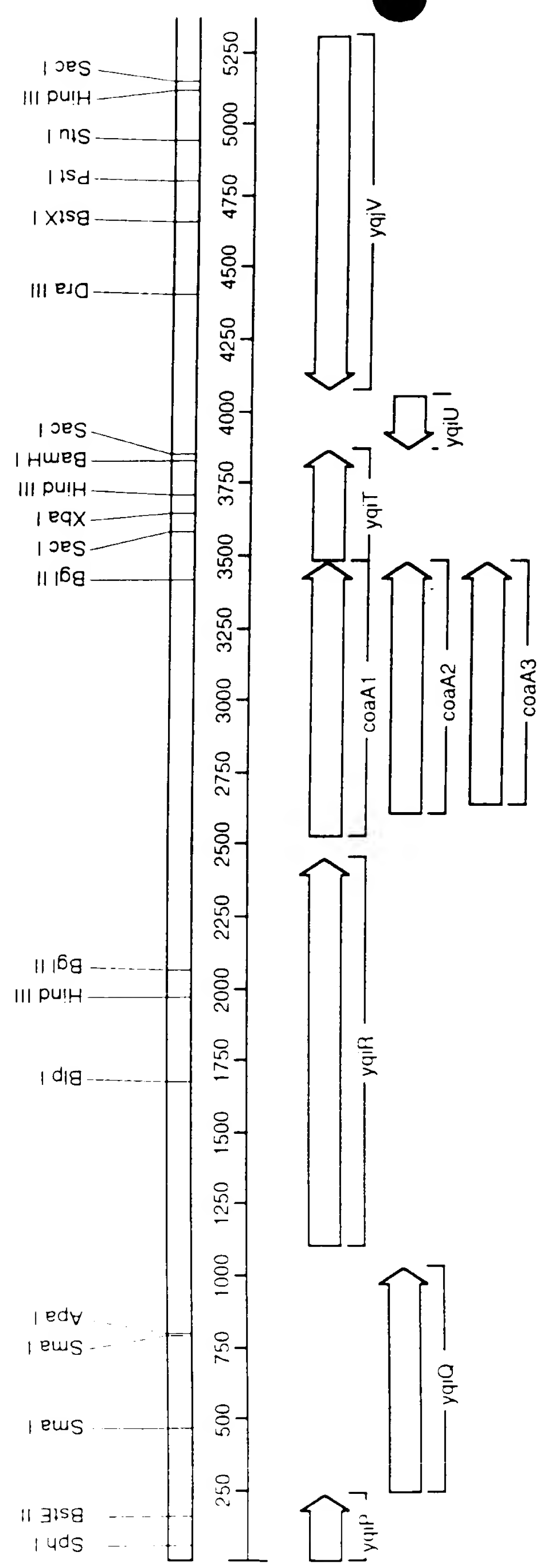


Figure 19 Structure of pAN281, a plasmid for expressing *B. subtilis* *coaA* after integration at the *bpr* locus. pAN282 and pAN283 have similar structures.

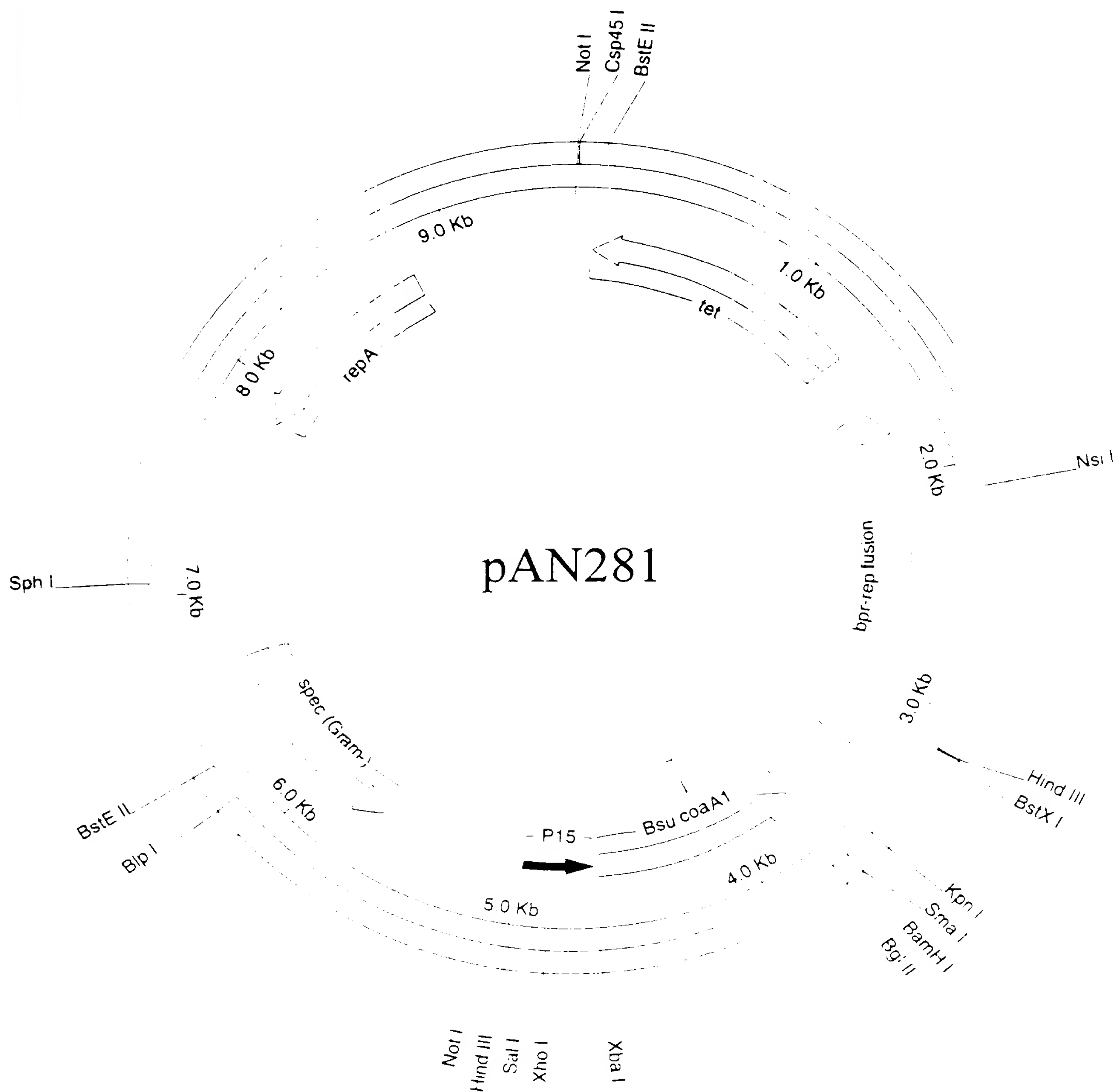


FIG. 20A

# THE HISTORY OF THE UNITED STATES

[illegible][illegible]

# THE HISTORY OF THE UNITED STATES

[illegible]

1. *What is the purpose of the study?*  
 2. *What are the research questions or hypotheses?*  
 3. *What is the study design?*  
 4. *What is the sample size and how was it selected?*  
 5. *What are the variables being measured?*  
 6. *What are the data collection methods?*  
 7. *What are the results of the study?*  
 8. *What are the conclusions of the study?*  
 9. *What are the limitations of the study?*  
 10. *What are the implications of the study?*

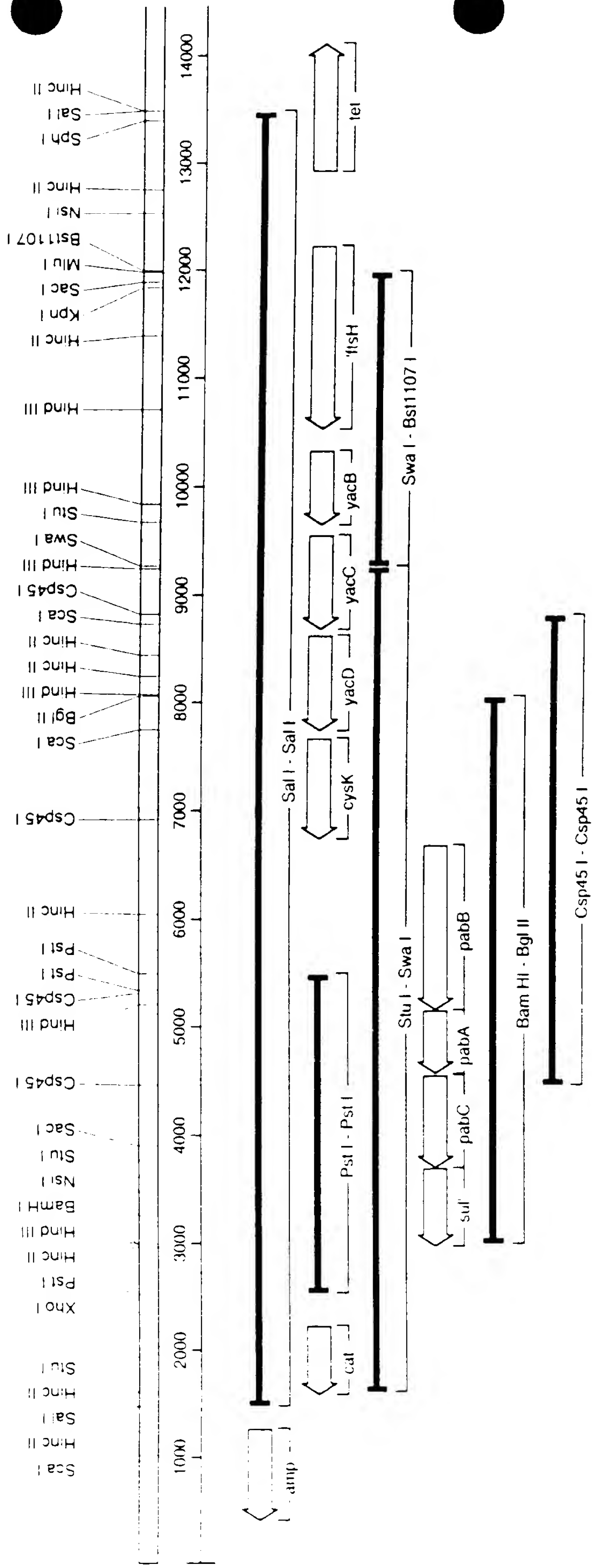
1. *Chlorophyll a* and *Chlorophyll b* were determined using a spectrophotometer (Shimadzu UV-1601) at 663 nm and 646 nm, respectively. The concentration of chlorophyll was calculated using the following formula:  $\text{Chlorophyll } a \text{ (mg/L)} = 12.7 \times \text{Absorbance at 663 nm}$  and  $\text{Chlorophyll } b \text{ (mg/L)} = 22.9 \times \text{Absorbance at 646 nm}$ .

# 2025年1月1日

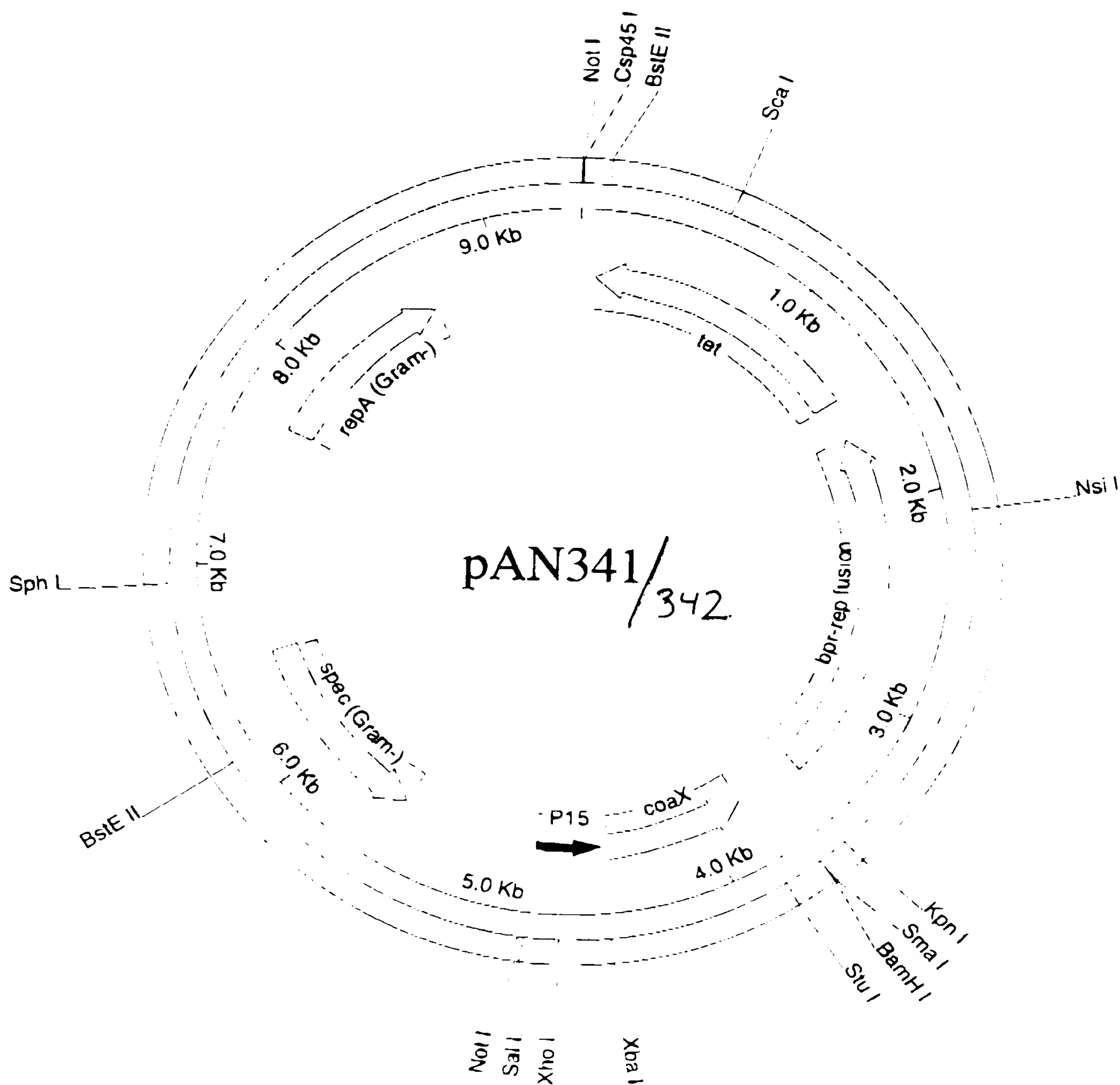
[illegible][illegible][illegible]



Figure 21



**Figure 22** Structure of pAN341 and pAN342, two independent PCR-derived clones of *yacB* (renamed *coaX*).





## FIG. 23A

[illegible]

absorption of the complexly with to protein  
 before to form to fourfold

Gen. 1: B.4144110, GenZ13EJH6-9	253	ad	Gen. 8: sp1051477, B.burgdorferi	262	ad
Gen. 2: B.414521426, 116, vulgare	212	ad	Gen. 9: sp1774049, Synchocystis	277	ad
Gen. 3: B.4145947, T.aurum	246	ad	Gen. 11: sp1025333, H. pylori	285	ad
Gen. 4: B.4146341, A. californica	265	ad	Gen. 11: sp1067003, A. acidicus	285	ad
Gen. 5: sp1243381B, portuensis	267	ad	Gen. 12: sp109R2541D, radiodurans	286	ad
Gen. 6: sp1000023, R. vulgare	272	ad	Gen. 13: WIT.RK03301, A. stratum	287	ad
Gen. 7: sp1000046, R. rubrum	273	ad	Gen. 14: WIT.RK03347, R. vesulatus	287	ad

[illegible]





FIG. 23D

[illegible]

VERBODEN  
VERVOER  
VERGEEFDE  
VERKONING  
WASROEKE  
OFDO  
ARUMPTALIAVENSON  
ONSIDERT  
LYLTH  
RHODRI  
LKKAOLAK  
WAOMPA

1. *Chrysomelidae*. *Chrysomelidae*.  
 2. *Chrysomelidae*. *Chrysomelidae*.  
 3. *Chrysomelidae*. *Chrysomelidae*.  
 4. *Chrysomelidae*. *Chrysomelidae*.  
 5. *Chrysomelidae*. *Chrysomelidae*.  
 6. *Chrysomelidae*. *Chrysomelidae*.  
 7. *Chrysomelidae*. *Chrysomelidae*.  
 8. *Chrysomelidae*. *Chrysomelidae*.  
 9. *Chrysomelidae*. *Chrysomelidae*.  
 10. *Chrysomelidae*. *Chrysomelidae*.

1. The first step is to identify the problem or goal. This involves understanding the current situation and what needs to be achieved.

2. Next, it's important to gather information and resources. This could involve research, consulting experts, or identifying the people and tools needed.

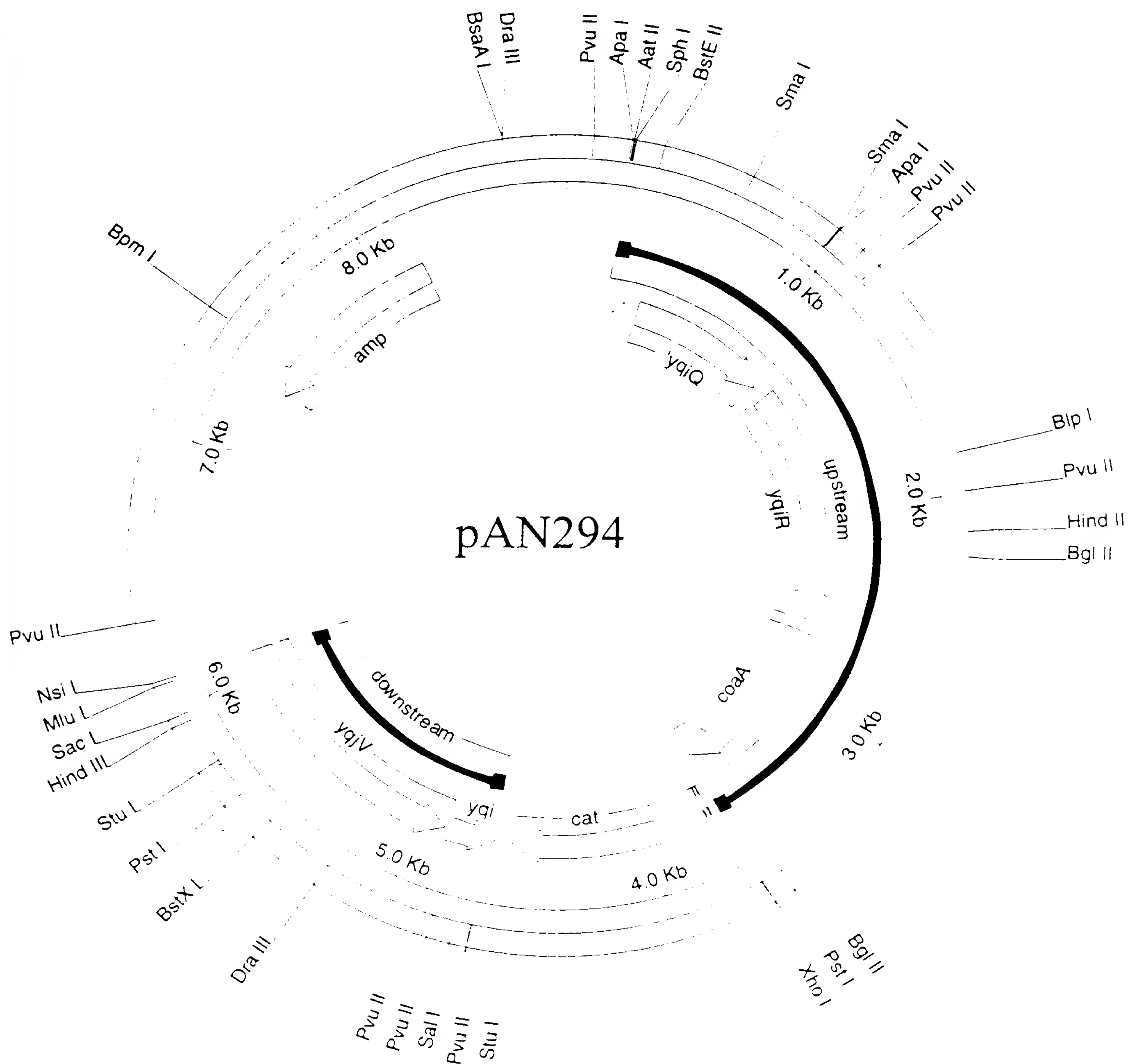
3. Once you have the information, you can start to develop a plan. This plan should outline the steps you will take to achieve your goal.

4. After the plan is developed, it's time to implement it. This involves putting the plan into action and making adjustments as needed.

5. Finally, it's important to evaluate the results. This involves checking to see if the goal has been achieved and if the process was effective.



*Figure 25 Structure of pAN294, a plasmid for integrating mutagenized B. subtilis coaA at its native locus.*



**Figure 26** Structure of pAN336, a plasmid designed to delete *B. subtilis* *coaX* from the chromosome and replace it with a kanamycin resistance gene.

